Fluid Biomarkers in Alzheimer Disease

Kaj Blennow1, Henrik Zetterberg1, and Anne M. Fagan2

1Clinical Neurochemistry Laboratory, Institute of Neuroscience and Physiology, Department of Psychiatry and Neurochemistry, The Sahlgrenska Academy at University of Gothenburg, Sahlgrenska University Hospital, Mölndal, SE-431 80 Mölndal, Sweden
2Department of Neurology, The Charles F. and Joanne Knight Alzheimer’s Disease Research Center, Hope Center for Neurological Disorders, Washington University School of Medicine, St. Louis, Missouri 63110

Correspondence: kaj.blennow@neuro.gu.se

Research progress has provided detailed understanding of the molecular pathogenesis of Alzheimer disease (AD). This knowledge has been translated into new drug candidates with putative disease-modifying effects, which are now being tested in clinical trials. The promise of effective therapy has created a great need for biomarkers able to detect AD in the predementia phase, because drugs will probably be effective only if neurodegeneration is not too advanced. In this chapter, cerebrospinal fluid (CSF) and plasma biomarkers are reviewed. The core CSF biomarkers total tau (T-tau), phosphorylated tau (P-tau) and the 42 amino acid form of β-amyloid (Aβ42) reflect AD pathology, and have high diagnostic accuracy to diagnose AD with dementia and prodromal AD in mild cognitive impairment cases. The rationale for the use of CSF biomarkers to identify and monitor the mechanism of action of new drug candidates is also outlined in this chapter.

In 1906, Alois Alzheimer presented the first case of the disease that was to bear his name, Alzheimer disease (AD). Alzheimer described the “miliary bodies” (plaques) and “dense bundles of fibrils” (tangles) which we today know are the hallmarks of the disease. In 1985, researchers succeeded in purifying plaque cores and amyloid angiopathy, and the 4 kD β-amyloid (Aβ) peptide was identified as the main component (Glenner and Wong 1984; Masters et al. 1985). This breakthrough paved the way for the cloning of the amyloid precursor protein (APP) gene (Kang et al. 1987). Almost at the same time, it was shown that tangles are composed of abnormally hyperphosphorylated tau protein (Grundke-Iqbal et al. 1986). These important achievements marked the start of modern AD research.

Today, detailed knowledge is available about APP metabolism and Aβ generation and on tau protein homeostasis. Largely based on the mutations found in familial AD (FAD), Aβ has been proposed as the driving force in the disease process. In line with this, the “amyloid cascade hypothesis” for AD (Hardy and Selkoe 2002) posits that an imbalance between the production and clearance of Aβ is the initiating event in disease pathogenesis, ultimately leading to neuronal degeneration and dementia. This research progress has been translated into
novel treatment strategies with disease-modifying potential. A large number of anti-Ab drug candidates, such as those used in Ab immunotherapy, secretase inhibitors, and Ab aggregation inhibitors, are in various phases of clinical treatment trials (Blennow et al. 2006). It should be noted, however, that the amyloid cascade hypothesis has not been proven with certainty in late-onset AD, the most common form of the disease.

Disease-modifying drugs will probably be most effective in the earlier stages of the disease, before plaque and tangle load and neurodegeneration become too severe (Das et al. 2001; Levites et al. 2006; Garcia-Alloza et al. 2009). Thus, these treatments should be administered in the predementia stage, or even in presymptomatic individuals. Further, in order for treatments to be labeled as “disease-modifying,” they must show a beneficial effect on cognition, as well as evidence that the drug does indeed affect the central disease processes and hallmark neuropathology (Siemers 2009). These challenges have created a need for biomarkers that reflect core elements of the disease process, to serve as diagnostic aids and as tools to identify and monitor the biochemical mechanism of action of the drug. In this chapter, we review the development of candidate biomarkers of AD from cerebrospinal fluid (CSF) and plasma. We focus on established biomarkers, those that have been evaluated in several studies by different research groups, and we discuss their implementation in clinical routine and their potential role in clinical trials.

FLUID BIOMARKERS AND THE BRAIN

Biomarkers are objective measures of a biological or pathogenic process that can be used to evaluate disease risk or prognosis, to guide clinical diagnosis, or to monitor therapeutic interventions (Blennow et al. 2010). The CSF is in direct contact with the extracellular space of the brain, and biochemical changes in the brain are therefore reflected in the CSF. The CSF is thus the optimal source for AD biomarkers.

Since Ab42 and tau have been shown to be the primary protein components of amyloid plaques and neurofibrillary tangles, respectively, the levels of these proteins in CSF have been assessed as potential biomarkers of these pathologic features (Fig. 1). Levels of Ab42 in postmortem ventricular CSF have been shown to correlate with plaque load at autopsy (Strozky et al. 2003). Similar results have been reported in antemortem lumbar CSF; with low levels correlating with postmortem plaque load (Tapiola et al. 2009). The development of Ab ligands suitable for positron emission tomography (PET) has enabled direct visualization of fibrillar Ab load in the brain in living individuals. Studies have consistently found a relationship between in vivo amyloid load as assessed by Pittsburgh Compound B (PIB)-PET binding and CSF Ab42, with higher 11C-PIB binding correlating with lower CSF Ab42 levels (Fagan et al. 2006; Forsberg et al. 2008; Grimmer et al. 2009; Tolboom et al. 2009). A similar relationship has been found between CSF Ab42 and binding of 18FFDDNP, a PET ligand believed to label both plaques and tangles (Tolboom et al. 2009). These results support the idea that CSF Ab42 is a measure of fibrillar Ab42 and plaque load in the brain. The most widely accepted explanation for the reduced CSF level of Ab42 in AD is that the aggregation of Ab into plaques (and thus retention in the brain parenchyma) results in less Ab being available to diffuse into the CSF.

Evidence that CSF total tau (T-tau) reflects the intensity of the neuronal and axonal damage and degeneration has come from several types of studies (Fig. 1). CSF T-tau increases markedly and transiently in acute disorders such as stroke and brain trauma, and the magnitude of the increase positively correlates with the size of the damaged tissue and negatively correlates with clinical outcome (Hesse et al. 2001; Ost et al. 2006; Zetterberg et al. 2006). The degree of increase in CSF T-tau in chronic neurodegenerative disorders is highest in disorders with the most rapid neuronal degeneration, such as Creutzfeldt–Jakob disease (Otto et al. 1997). High CSF T-tau is also associated with a faster progression from mild cognitive impairment (MCI) to AD (Blom et al. 2009), and a more rapid cognitive decline and higher mortality in...
AD cases (Samgard et al. 2009; Wallin et al. 2009). However, one study found that CSF T-tau correlates with postmortem tangle load (Tapiola et al. 2009), suggesting that the release of tau specifically from degenerating tangle-bearing neurons may contribute to the CSF level of T-tau. Consistent with this idea, binding of {\textsuperscript{18}F}DDNP, an agent that is reported to label both plaques and tangles, positively correlates with CSF T-tau levels (Tolboom et al. 2009).

It is logical to postulate that phosphorylated tau (P-tau) in CSF reflects the phosphorylation state of the tau protein in the central nervous system (CNS; Fig. 1). Positive correlations between CSF levels of P-tau_{181} and P-tau_{231} (tau phosphorylated at residues 181 and 231, respectively) and neocortical tangle pathology at autopsy have been reported (Buerger et al. 2006; Tapiola et al. 2009). High CSF P-tau_{181} is also associated with a faster progression from MCI to AD (Blom et al. 2009), and a more rapid cognitive decline in AD cases (Samgard et al. 2009), as well as those with very mild AD dementia (Snider et al. 2009). These findings support the hypothesis that the CSF level of P-tau reflects the phosphorylation state of tau and the formation of tangles in the brain.

**FLUID BIOMARKERS FOR AD DIAGNOSIS**

**β-Amyloid Isoforms**

The discovery that β-amyloid (Aβ) is produced during normal cell metabolism and is secreted into the CSF served as the basis for Aβ biomarker development (Seubert et al. 1992). The subsequent finding that Aβ_{42} is the most abundant species in plaques made it logical to develop assays for this Aβ isoform (Jarrett et al. 1993). CSF Aβ_{42} in AD is decreased to approximately 50% of control levels, as has been shown using several different enzyme-linked immunosorbent assay (ELISA) methods (Sunderland et al. 2003; Blennow 2004).
There are several isoforms of the tau protein in CSF, and the molecule has numerous phosphorylation sites (Portelius et al. 2008). The most commonly used ELISA method for T-tau is based on monoclonal antibodies that detect all isoforms of tau independently of phosphorylation state (Blennow et al. 1995). Numerous studies have used this assay, and consistently report a marked increase of CSF T-tau in AD to around 300% of control levels (Sunderland et al. 2003; Blennow 2004).

The most commonly used ELISA methods for P-tau in CSF use antibodies that are specific for phosphorylation at either threonine 181 (P-Tau181) or threonine 231 (P-Tau231; Kohnken et al. 2000; Vanmechelen et al. 2000). Studies using these assays have consistently found a marked increase in CSF P-tau in AD (Blennow 2004). Research that compared these P-tau assays directly found a very high correlation between the methods and similar diagnostic performances (Hampel et al. 2004).

Several studies have shown that the diagnostic accuracy for the combination of CSF T-tau, P-tau and Aβ42 is higher than for any biomarker alone (Galasko et al. 1998; Reimenschneider et al. 2002; Maddalena et al. 2003; Zetterberg et al. 2003; Hansson et al. 2006). A logical strategy, therefore, was to develop a multiparameter assay for simultaneous quantification of these CSF biomarkers, based on the Luminex™ xMAP technology (Olsson et al. 2005). This assay has been used in several recent large multicenter studies on CSF biomarkers, and its diagnostic performance has been good (Hansson et al. 2006; Lewczuk et al. 2008; Mattsson et al. 2009; Shaw et al. 2009). The measured values for CSF levels of the biomarkers differ between the Luminex technique and the ELISA methods (Olsson et al. 2005; Lewczuk et al. 2008). There are likely to be several reasons for this, including differences in the pairs of antibodies selected, the method for coupling antibodies to beads, the method of coating plates, differences in the calibrators, and differences in the incubation conditions. Correction factors have been used to convert results from one technique to the other, allowing the results to be compared (Olsson et al. 2005; Mattsson et al. 2009).

AD with Dementia

Numerous studies have found a marked increase in CSF T-tau and P-tau, together with a marked decrease in Aβ42, in AD cases with dementia. These measurements can be used to discriminate patients with AD from the nondemented aged with a sensitivity and specificity that both lie above 80% (Blennow and Hampel 2003; Blennow 2004). CSF levels of these markers are normal in several important differential diagnoses, such as depression and Parkinson disease (Blennow 2004). Combined analyses of these biomarkers give a better diagnostic performance than any biomarker alone (Galasko et al. 1998; Maddalena et al. 2003; Hansson et al. 2006; Mattsson et al. 2009). CSF P-tau, in particular, aids in the differentiation of AD from other dementias, such as frontotemporal dementia and Lewy body dementia (Hampel et al. 2004), but the diagnostic performance of CSF biomarkers to discriminate AD from other dementias is not optimal. There are several reasons for this. First, most studies of CSF biomarkers are based on clinically diagnosed cases, which introduces a relatively large percentage of misdiagnosis (Blennow 2005; Forman et al. 2006). Second, a significant percentage of the nondemented elderly have enough plaques and tangles to warrant a neuropathological diagnosis of AD (Snowdon 1997; Price and Morris 1999). Third, there is a large overlap in pathology between AD and other dementias, such as Lewy body dementia and vascular dementia (Jellinger 1996; Kotzbauer et al. 2001; Schneider et al. 2009). This overlap in pathology essentially precludes the possibility...
of finding any biomarkers that have close to 100% sensitivity and specificity for AD.

Autopsy-Verified AD

Several studies have examined the diagnostic performance of CSF biomarkers in patient series in which diagnosis can subsequently be confirmed by autopsy. CSF biomarkers have high sensitivity and specificity in discriminating AD from both the cognitively normal elderly and from patients with other dementias, such as frontotemporal dementia, Lewy body dementia and vascular dementia (Clark et al. 2003; Sunderland et al. 2003; Bian et al. 2008; Koopman et al. 2009; Shaw et al. 2009). CSF biomarkers have thus been validated in patient series with a neuropathological follow-up, showing similar or better discriminatory power than in patient series with clinical diagnoses only.

Prodromal AD

Cerebrospinal fluid biomarkers also have a high predictive value in identifying prodromal AD in MCI cases (Blennow and Hampel 2003). A recent study with an extended clinical follow-up period showed that the combination of all three core CSF biomarkers (T-tau, P-tau, and Aβ42) had a sensitivity of 95% for the identification of prodromal AD in MCI (Hansson et al. 2006). These CSF markers have also been shown to predict the rate of cognitive decline in patients with MCI/very mild AD dementia (Snider et al. 2009). A high predictive value has also been verified in large multicenter studies, including the ADNI study (Shaw et al. 2009), the DESCRIPA study (Visser et al. 2009), and the Swedish Brain Power project (Mattsson et al. 2009). These results demonstrate that CSF biomarkers may be valuable clinical diagnostic tools to identify MCI cases with prodromal AD.

Preclinical AD

Preclinical AD denotes cognitively normal individuals harboring early AD pathology, not severe enough to cause cognitive symptoms. Some studies have examined whether CSF biomarkers are useful in the preclinical stage to identify patients who will subsequently develop AD dementia. Two population-based studies found a significant reduction in CSF Aβ42 in cognitively normal elderly who later developed AD, whereas there was no significant change in CSF T-tau or P-tau (Skoog et al. 2003; Gustafson et al. 2007). A recent clinical study also found that CSF Aβ42, but not T-tau and P-tau, predicts cognitive decline in the healthy elderly (Stomrud et al. 2007). Four independent studies have identified the CSF tau/Aβ42 ratio (but not these markers individually) as a strong predictor of future cognitive decline (within a few years) in nondemented elders (Fagan et al. 2007; Li et al. 2007; Craig-Schapiro et al. 2010; Tarawneh et al. 2011), similar to its ability to predict AD dementia in MCI cohorts (Hansson et al. 2006). Asymptomatic carriers of FAD mutations also have low CSF Aβ42 (Moonis et al. 2005), and high T-tau and P-tau (Ringman et al. 2008). These results extend earlier animal data suggesting that the amyloidogenic process is upstream of tau pathology in AD (Gotz et al. 2001; Lewis et al. 2001). Consistent with this idea, two recent studies reported an association between low CSF Aβ42 levels and brain atrophy in cognitively normal elders (Fagan et al. 2009a; Fjell et al. 2010), whereas CSF tau and ptau181 levels were associated with atrophy in early-stage MCI/AD (Fagan et al. 2009a). These biomarker observations suggest that Aβ aggregation and deposition (as evidenced by reduced CSF Aβ42) are associated with brain atrophy in the preclinical phase of the disease, whereas changes in CSF tau and accelerated brain atrophy are later events in the disease that occur with or just prior to cognitive decline and subsequent clinical progression (Fig. 2). Additional studies are required to confirm these findings.

A recent large study showed that cognitively normal elderly who are positive for PIB-PET (indicating the presence of brain amyloid) have low levels of CSF Aβ42 (Fagan et al. 2009b), confirming results from an earlier, smaller study (Fagan et al. 2006). However, in this larger cohort, CSF Aβ42 was found to also be low in a small subset of PIB-negative
individuals (Fagan et al. 2009b). This finding suggests that low CSF Aβ42 may serve as a harbinger of future amyloid deposition in the preclinical period. Longitudinal PIB follow-up in these individuals will be required to test this hypothesis. Alternatively, low CSF Aβ42 in the absence of PIB positivity may be an indicator of Aβ aggregation in diffuse (PIB-negative) plaques or the accumulation of oligomeric species of Aβ within brain parenchyma prior to substantial fibrillar (PIB-positive) Aβ deposition, or may simply reflect the low end of the normal spectrum of CSF Aβ42 levels. In support of the first alternative hypothesis, one of the PIB-negative individuals with low CSF Aβ42 came to autopsy and was found to have widespread diffuse, but minimal fibrillar, plaque deposits, suggesting that low CSF Aβ42 may mark diffuse plaques in addition to fibrillar plaques (Cairns et al. 2009). Regardless of the underlying biological mechanism(s), these results suggest that CSF Aβ42 is a marker of AD plaque pathology very early in the disease process (prior to cognitive symptoms). However, it remains to be determined whether CSF Aβ42 levels will allow the prediction of prodromal AD in individual cases. Also, although these data offer important insights into the normal pathophysiology of the disease, the use of biomarkers to predict AD in the asymptomatic elderly is not warranted until registered drugs are available that offer a distinct disease-modifying effect combined with few side-effects. However, such biomarkers may be very useful in the immediate future for the design and evaluation of prevention trials by allowing one to enroll individuals who are still cognitively normal but are in the preclinical stage of the disease.

Figure 2. Hypothetical model of the temporal evolution of biomarkers for Alzheimer disease (AD) (top) in relation to pathogenic processes in the brain (middle) and clinical stage of the disease (bottom).
and, importantly, are within a few years of developing cognitive symptoms.

**NOVEL FLUID BIOMARKERS FOR AD**

There are numerous publications describing candidate CSF biomarkers other than Aβ and tau, but initial promising results have most often not been reproduced (Table 1). Here, we review novel biomarkers that have shown promise in two independent studies, and shown a reasonable sensitivity and specificity for AD. We also discuss some candidate biomarkers specifically related to Aβ and APP metabolism.

**sAPPβ and sAPPα**

During APP processing, the large amino-terminal domains of APP, sAPPα, and sAPPβ, are secreted into the extracellular space and eventually reach the CSF. In sporadic AD and MCI, CSF levels of both sAPPα and sAPPβ are unaltered or slightly increased (Olsson et al. 2003; Zetterberg et al. 2008; Lewczuk et al. 2010). Although there is no consistent change in sAPP levels in AD, these CSF biomarkers may be valuable tools in treatment trials to monitor an effect on APP processing.

**BACE1**

The major β-secretase responsible for Aβ generation is β-site APP-cleaving enzyme 1 (BACE1). BACE1 expression and enzymatic activity are both increased in postmortem brains of patients with AD (Fukumoto et al. 2002; Yang et al. 2003). BACE1 can be measured in CSF, and its concentration and activity increase in AD, preferentially in MCI cases with prodromal AD (Holsinger et al. 2004; Zhong et al. 2007; Zetterberg et al. 2008). These results suggest that up-regulation of BACE1 is an early pathogenic event in AD.

**Aβ Oligomers**

Aggregation of soluble Aβ to form insoluble fibrillar aggregates in plaques has long been regarded as the central pathogenic event in AD. However, recent results suggest that soluble Aβ oligomers inhibit long-term potentiation, the proposed biological substrate of memory, thereby playing a role in AD pathogenesis (Walsh and Selkoe 2007). Measurement of Aβ oligomers in CSF may thus be an important core biomarker for AD. Some preliminary studies on Aβ oligomers in CSF have been published. Using monoclonal antibodies for Aβ oligomers in an assay with PCR-based signal amplification, one study reported a marked increase in AD autopsy CSF (Georganopoulou et al. 2005). Flow cytometric results also suggest that Aβ oligomers are present in CSF, but no information on the diagnostic utility of this assay has been presented (Santos et al. 2007). A weak band migrating at the size expected for Aβ dimers appears in CSF immunoprecipitation experiments, utilizing an anti-Aβ antibody followed by SDS–PAGE and immunoblotting, but there is no clear correlation with AD (Klyubin et al. 2008). Immunoassays in which a monoclonal antibody is used for capture, and the same, biotinylated, antibody is used for detection, may be used to quantify protein aggregates, because monomers will not be detected since the epitope is already occupied by the capture antibody. Using ELISA methods based on this principle, higher CSF levels of Aβ oligomers were found in AD (Fukumoto et al. 2010). These promising results call for further studies, and need to be replicated in larger independent patient materials. It seems clear that although Aβ oligomers are an attractive AD biomarker candidate, the level of such oligomers is very low compared with that of Aβ monomers. Further, the identity of signals measured using different techniques must be verified by mass spectrometry.

**Other Aβ Isoforms**

Aβ40 is the most abundant Aβ isoform in CSF (Portelius et al. 2006a). Although there is no major change in the level of CSF Aβ40 in AD, and Aβ40 levels do not correlate with amyloid load as evidenced by PIB binding (Fagan et al. 2006, 2009b), there is a marked decrease in the ratio of CSF Aβ42/Aβ40 in AD and MCI, which is more pronounced than the reduction in CSF...
<table>
<thead>
<tr>
<th>Pathogenic process</th>
<th>Biomarker</th>
<th>Methodology</th>
<th>Change in AD</th>
<th>Stage of evaluation</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>APP/Aβ metabolism and plaque pathology</td>
<td>CSF Aβ42 and Aβ40</td>
<td>Commercially available in several different immunoassay formats. Assay characteristics and confounding factors well established</td>
<td>Approximately 50% reduction in CSF Aβ42 in AD with dementia and prodromal AD. CSF Aβ42/Aβ40 ratio may give slightly higher accuracy than Aβ42 alone</td>
<td>Consistent results from numerous publications</td>
<td>CSF Aβ42 is the central CSF biomarker for Aβ metabolism. CSF Aβ42 correlates with amyloid load measured by PIB-PET.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Plasma Aβ42 and Aβ40</td>
<td>Commercially available in several different immunoassay formats</td>
<td>No consistent change in AD. Large overlap with healthy elderly</td>
<td>Consistent results from numerous publications</td>
<td>Plasma Aβ42 and Aβ40 has no diagnostic value for AD. May be valuable in clinical trials.</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CSF Aβ16</td>
<td>Immunoprecipitation combined with MALDI-TOF mass spectrometry</td>
<td>Increase in CSF Aβ16 in AD</td>
<td>Preliminary data. Needs verification using standard immunoassays</td>
<td>Analysis of the Aβ isoform pattern, including Aβ16, may be of value in clinical trials on secretase inhibitors.</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CSF Aβ oligomers</td>
<td>Specialized techniques, e.g., Bio-barcode PCR assay, western blot, or high-sensitivity ELISA</td>
<td>Increased level or increased frequency of Aβ oligomers in AD</td>
<td>Preliminary data. Needs verification using standard immunoassays</td>
<td>The nature (dimers, trimers, dodecamers, high MW species) of Aβ oligomers in CSF has to be determined.</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CSF APP isoforms (sAPPα, sAPPβ)</td>
<td>Commercial assays available</td>
<td>No change or slight increase in AD with large overlap with healthy elderly</td>
<td>Needs further evaluation</td>
<td>APP isoforms are not diagnostically useful, but may be valuable in clinical trials on, e.g., BACE1 inhibitors.</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CSF BACE1 activity</td>
<td>Different research assays used in different publications</td>
<td>Increase in AD with dementia and prodromal AD</td>
<td>Data based on publications using different methods</td>
<td>The diagnostic value needs further evaluation. BACE1 activity may be useful in clinical trials on, e.g., BACE1 inhibitors.</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Brain Aβ turnover</td>
<td>Infusion of labeled leucine and continuous CSF sampling; immunoprecipitation, tryptic digestion and mass spectrometry measurement of total Aβ</td>
<td>First study shows a decreased Aβ turnover</td>
<td>Needs further evaluation</td>
<td>May be valuable to gauge Aβ production and clearance in clinical drug trials.</td>
<td></td>
</tr>
</tbody>
</table>

Continued
<table>
<thead>
<tr>
<th>Pathogenic process</th>
<th>Biomarker</th>
<th>Methodology</th>
<th>Change in AD</th>
<th>Stage of evaluation</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tau phosphorylation and tangle pathology</td>
<td>CSF P-tau181</td>
<td>Commercially available in different immunoassay formats. Assay characteristics and confounding factors well established</td>
<td>Increase in AD with dementia and prodromal AD</td>
<td>Consistent results from numerous publications</td>
<td>CSF P-tau is the central CSF biomarker for tau phosphorylation state. May be valuable as a downstream biomarker in anti-Aβ treatment trials</td>
</tr>
<tr>
<td></td>
<td>CSF P-tau231</td>
<td>Commercial assay not available</td>
<td>Increase in AD with dementia and prodromal AD</td>
<td>Consistent results from numerous publications</td>
<td>CSF P-tau is the central CSF biomarker for tau phosphorylation state. May be valuable as a downstream biomarker in anti-Aβ treatment trials</td>
</tr>
<tr>
<td>Neuronal and axonal degeneration</td>
<td>CSF T-tau</td>
<td>Commercially available in different immunoassay formats. Assay characteristics and confounding factors well established</td>
<td>Increase in AD with dementia and prodromal AD</td>
<td>Consistent results from numerous publications</td>
<td>CSF T-tau is the central CSF biomarker to monitor the intensity of neuronal and axonal degeneration in treatment trials</td>
</tr>
<tr>
<td></td>
<td>CSF VILIP-1</td>
<td>Single (research) assay</td>
<td>Increase in AD</td>
<td>Needs further evaluation</td>
<td>CSF VILIP-1 levels correlate with CSF T-tau. May be a valuable complementary biomarker for axonal degeneration</td>
</tr>
<tr>
<td></td>
<td>CSF NF proteins</td>
<td>Different (research) assays using in different publications</td>
<td>Normal in AD. High CSF NF proteins in disorders with subcortical pathology, e.g., VaD and NPH, and in FTD</td>
<td>Consistent results from numerous publications</td>
<td>CSF NF proteins may be valuable to differentiate AD from frontotemporal dementia and subcortical dementia disorders</td>
</tr>
</tbody>
</table>

Abbreviations: AD, Alzheimer disease; BACE1, β-site APP-cleaving enzyme 1; CSF, cerebrospinal fluid; FTD, frontotemporal dementia; NF, neurofilament; NPH, normal pressure hydrocephalus; PET, positron emission tomography; PIB, Pittsburgh compound B; P-tau, phosphorylated tau; T-tau, total tau; VaD, vascular dementia; VILIP-1, visinin-like protein 1.
Aβ42 alone (Mehta et al. 2000; Hansson et al. 2007a). Apart from Aβ42 and Aβ40, other carboxy-terminally truncated Aβ peptides can be identified in CSF, including Aβ37, Aβ38, and Aβ39 (Lewczuk et al. 2003). The CSF level of Aβ38 has been reported to be higher in AD, together with a decrease in the level of Aβ42 (Lewczuk et al. 2003; Schoonenboom et al. 2005), suggesting that the Aβ42/Aβ38 ratio may be used to improve diagnostic accuracy. The ratio of Aβ38/Aβ42 has also been shown to positively correlate with PIB binding in nondemented cohorts (Fagan et al. 2009b), with an association that is slightly stronger than CSF Aβ42 alone.

Several shorter carboxy-terminally truncated Aβ isoforms in CSF have also been identified and quantified by immunoprecipitation with an anti-Aβ monoclonal antibody and Matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry (Portelius et al. 2006a). An increase in CSF Aβ1–16 is found in AD together with the expected decrease in Aβ1–42 (Portelius et al. 2006b, 2010). Data from experimental studies show that the shorter Aβ isoforms Aβ1–14, Aβ1–15, and Aβ1–16 are produced by a novel pathway for APP processing involving the concerted action of β- and α-secretase, whereas the longer isoforms, from Aβ1–17 and up to Aβ1–42, are produced in the γ-secretase pathway (Portelius et al. 2009).

**Neuronal and Synaptic Degeneration**

Neuronal and synaptic proteins may prove valuable as CSF biomarkers because they provide information about cognitive function and disease progression. For example, visinin-like protein 1 (VILIP-1) is a highly expressed neuronal calcium sensor protein that was identified by gene array analyses designed to search for brain-specific protein biomarkers (Laterza et al. 2006). CSF VILIP-1 increased markedly in AD in a clinical study, with a diagnostic performance similar to that of CSF tau and Aβ (Lee et al. 2008). CSF levels of VILIP-1 were higher in APOE ε4-positive cases, and correlated with Mini-Mental State Examination (MMSE) scores. A recent study of VILIP-1 showed that the ratio of VILIP-1/Aβ42 was as good or better than tau/Aβ42 in predicting progression from cognitively normal to very mild dementia (Tarawneh et al. 2011).

Neurofilament (NF) proteins are structural components of the neuronal axons. The expression of such proteins is particularly high in large myelinated axons (Friede and Samorajski 1970). Accordingly, high CSF levels of NF proteins are found in disorders with subcortical pathology, such as vascular dementia and normal-pressure hydrocephalus (Sjogren et al. 2001a; Agren-Wilson et al. 2007). CSF levels are also high in frontotemporal dementia, whereas they are normal in most cases of AD (Sjogren et al. 2000). CSF NF proteins may thus be valuable for differentiation between AD, frontotemporal dementia, and subcortical dementia disorders.

Clinicopathologic studies have demonstrated that synaptic density is the variable that best correlates with cognitive performance (Terry et al. 1991). Thus, one might predict that synaptic proteins would be the class of biomarkers most tightly linked to cognition. Several pre- and postsynaptic proteins have been identified in CSF using a procedure based on protein precipitation followed by liquid-phase isoelectric focusing and western blotting. These proteins include rab3a, synaptotagmin, growth-associated protein (GAP-43), synaptosomal-associated protein (SNAP-25), and neurogranin (Davidsson et al. 1999). An immunoassay for GAP-43 showed that CSF levels are higher in AD patients than in controls and patients with frontotemporal dementia (Sjogren et al. 2001b). There are also positive correlations between CSF GAP-43 and T-tau, supporting the idea that both biomarkers reflect axonal and synaptic degeneration.

**Inflammation and Oxidative Stress**

Neuroinflammation, in the way of glial activation (especially in the vicinity of amyloid plaques), is a robust but nonspecific feature of AD. A number of reports published in the 1990s and early 2000s describe alterations in the levels
of various inflammatory and signaling molecules, as well as markers of oxidative stress (e.g., α1-antichymotrypsin, isoprostane, the interleukins, TNFα, interferon-γ, complement C1q, and TGF-β) in AD CSF (Zetterberg et al. 2004; Craig-Schapiro et al. 2009). However, results have been very inconsistent, probably owing to methodological differences (e.g., in the procedures for CSF collection and processing, assay differences, and criteria used for subject ascertainment), prevalence of comorbidities in the studied cohorts, and methods of diagnosis. Unbiased proteomics methods have more recently been used to identify molecules that differ between AD and control CSF (and serum and plasma). These studies have consistently identified a plethora of inflammatory markers that differ in abundance between clinical groups (Castano et al. 2006; Finehout et al. 2007). However, even in these unbiased screens, the direction of reported difference in abundance has not been consistent. Despite this, one astrocyte marker, YKL-40, discovered in an unbiased proteomic screen, has recently been validated in a large cohort of cognitively normal and AD subjects to be increased in AD and to predict clinical worsening from cognitively normal to very mild dementia (Craig-Shapiro et al. 2010). The recent availability of commercial multiplexed assays should permit analysis of a large panel of inflammatory and signaling molecules in large-scale studies. It is conceivable (and probable) that adding markers of neuroinflammation to the other CSF markers (such as Aβ42, tau, and P-tau) will further strengthen diagnostic and prognostic capability (Hu et al. 2010).

AD pathogenesis also includes free radical–mediated injury to neurons. Lipid peroxidation is an important consequence of such damage, and it generates many products, including F2-isoprostanes. These molecules may serve as biomarkers for this pathogenic process. CSF F2-isoprostane levels have been reported to be increased in AD (Montine et al. 2007). Recent studies also show an increase in F2-isoprostanes in MCI cases with prodromal AD (Brys et al. 2009a) and in asymptomatic carriers of FAD mutations (Ringman et al. 2008). In contrast, studies on F2-isoprostanes in plasma have reported conflicting results, probably because the contribution of brain-derived F2-isoprostanes to plasma is clouded by the much larger contribution of peripherally derived F2-isoprostanes (Montine et al. 2007).

**SILK Technology**

Recently a new in vivo technique, known as Stable Isotope Labeling Kinetics (SILK), has been developed to measure the production and clearance rates of CNS proteins in humans (Figure 3). In this technique, a stable (nonradioactive) isotope–labeled amino acid (e.g., 13C6-leucine) is administered intravenously and becomes incorporated into newly synthesized proteins. CSF (and plasma) can then be sampled over time via intrathecal and intravenous catheters, respectively. Using mass spectrometry to compare the amounts of labeled versus unlabeled proteins over time, very precise synthesis, clearance, and dose–response curves can be developed. This technique was first applied to determine the synthesis and clearance rates of Aβ in the CNS (Bateman et al. 2006). The fractional production and clearance rates of Aβ in vivo was found to be extremely rapid (7.6% per hr and 8.3% per hr, respectively), with absolute concentrations in the CSF varying widely between sampling times. This technique was used more recently in a randomized, double-blind, placebo-controlled study to demonstrate the pharmacokinetic/pharmacodynamic relationship between an Aβ synthesis inhibitor and the absolute rate of CNS Aβ synthesis (Bateman et al. 2009). In addition, this method identified slower fractional Aβ clearance with no change in fractional Aβ synthesis in late-onset AD versus age-matched controls (Mawuenyega et al. 2010). Since this technique automatically labels all newly synthesized proteins, its potential lies in the fact that it allows for the evaluation of other proteins relevant to AD, other neurodegenerative diseases, and the metabolism of multiple biomarkers simultaneously. As such, this technology may uncover robust fluid biomarkers that will be useful for assessing disease risk, improving AD diagnosis and prognosis.
tracking disease progression, and evaluating treatment efficacy.

**Plasma Biomarkers**

Efforts to find reliable biomarkers for AD in peripheral blood have had little success. Several candidate blood biomarkers have been proposed, but have been difficult to verify in independent studies. In this review we focus on plasma Aβ, which is the most extensively examined peripheral biomarker for AD. We also review some explorative pilot studies with promising results.

Many studies have examined plasma Aβ as a biomarker for AD, but the findings are contradictory. Some groups report slightly higher plasma levels of either Aβ42 or Aβ40 in AD, although with a broad overlap between patients and controls, whereas most studies find no change (Irizarry 2004). Studies examining the value of plasma Aβ in predicting AD in the cognitively normal elderly also show a very broad overlap in plasma Aβ42 and Aβ40 levels. Some studies report that high plasma Aβ42, or a high Aβ42/Aβ40 ratio, is a risk indicator for future AD, whereas others report the opposite (Mayeux et al. 2003; Pomara et al. 2005; van Oijen et al. 2006; Graff-Radford et al. 2007). These discouraging results are probably due to the fact that the majority of plasma Aβ is derived from peripheral tissues, and does not reflect brain Aβ turnover or metabolism (Mehta et al. 2000). This is consistent with a lack of correlation between plasma Aβ species and brain amyloid load as determined by PIB binding (Fagan et al. 2006, 2009b). It is possible, however, that the hydrophobic nature of Aβ causes it to bind to plasma proteins, which may result in epitope masking and other analytical interferences (Kuo et al. 1999).

There are some recent reports that present promising novel blood biomarkers for AD. Combined multivariate analysis of 18 plasma signaling and inflammatory proteins was found to identify AD patients and predict future AD,
with high accuracy, in MCI patients (Ray et al. 2007). This protein panel was identified after screening a large number of known proteins using a filter-based protein array. Further independent studies are needed to verify if this panel is the optimal combination of plasma biomarkers, as well as to determine their diagnostic value. Another study using explorative proteomics technology identified AD-associated changes in the plasma levels of complement factor H and α2-macroglobulin (Hye et al. 2006). This finding was replicated using semiquantitative immunoblotting techniques. A significant change was also reported in the ratio of the microcirculation regulating factor midregional pro-atrial natriuretic peptide to carboxy-terminal endothelin-1 precursor fragment, both of which regulate microcirculation, in plasma from AD patients (Buerger et al. 2009). If replicated in independent studies using immunosay techniques suitable for routine diagnostic laboratories, these types of plasma protein panels may serve as useful screening tests for AD.

FLUID BIOMARKERS IN CLINICAL TRIALS

In addition to their potential as tools for clinical diagnosis, CSF biomarkers may be valuable in drug development in at least four different ways. These uses are as diagnostic markers for the enrichment of AD cases, for patient stratification, as safety markers, and as detectors and monitors of biochemical drug effects (Table 2).

Improved Diagnosis/Enrichment of AD Cases

Diagnosing early AD is a great challenge for clinicians because MCI cases only have a mild disturbance in episodic memory and executive dysfunction, whereas other specific symptoms are lacking or are vague and indistinct. The only clinical method available to determine which MCI patients have prodromal AD is an extended follow-up period. Indeed, even at specialized academic centers, the accuracy of the clinical diagnosis of AD in cases that have been followed clinically for several years can be relatively low, with sensitivity and specificity figures of 70–80% (Knopman et al. 2001). These figures are considerably lower in patients with early AD (Visser et al. 2005) and in primary care settings (Ganguli et al. 2004). Assessment of cognitive changes within individuals over a number of years (as determined with semistructured interviews with the patient and a reliable collateral source who knows the patient well, such as a spouse or adult child), can increase diagnostic accuracy (Storandt et al. 2006). However, this laborious approach is probably not feasible in primary care settings.

Clinical MCI trials of cholinesterase inhibitors, in which having a reduced conversion rate to AD was used as a clinical endpoint, have failed to find any significant benefit of the drugs (Raschetti et al. 2007). These trials have recruited unselected MCI cases, meaning that approximately half of the cases do not have prodromal AD, and thus will not convert. This may have adversely affected the possibility of detecting any positive clinical effect of the drug (Cummings et al. 2007). Addition of positive CSF biomarkers as inclusion criteria in MCI and even prevention trials will increase the proportion of subjects with underlying AD pathology and thereby increase the possibility of detecting a positive effect of the drug.

Stratification of AD Cases Based on Biomarker Data

It is well established that AD is a heterogeneous disorder, at both the clinical and neuropathological levels (Blennow et al. 2006). It is quite possible that the effects of proposed disease-modifying drugs will differ between subgroups of AD patients with respect to degree of plaque and tangle pathology (as evidenced by biomarkers) or genetic determinants. As an example, passive immunization was reported to differ both in treatment effect and side-effects between APOE ε4 carriers and noncarriers (Salloway et al. 2009).

Since CSF biomarkers reflect the central pathogenic processes in AD, they may be used in post-hoc analyses to stratify the patient cohort in clinical trials. For example, one could postulate that a patient subgroup with a certain biomarker trait, such as low CSF Aβ42...
indicating plaque pathology, will show a better effect of anti-\(\mathrm{A}\beta\) disease-modifying drugs than a subgroup with normal CSF \(\mathrm{A}\beta 42\) levels.

### Safety Monitoring

Trials of the new type of disease-modifying treatments have been hampered by side-effects including meningoencephalitis, which was observed in a subset of cases in the AN-1792 trial on active \(\mathrm{A}\beta\) immunotherapy, and vasogenic edema, which was observed in the AAB-001 trial of passive immunotherapy (Orgogozo et al. 2003; Salloway et al. 2009). CSF analysis is the standard method to diagnose both encephalitis and blood–brain barrier damage associated with disorders that cause edema (Tibbling et al. 1977; Andersson et al. 1994). It is also possible to use the baseline CSF sample to initially identify and exclude cases with chronic infectious or

### Table 2. Use of fluid biomarkers in Alzheimer disease clinical trials

<table>
<thead>
<tr>
<th>Application</th>
<th>Rationale</th>
<th>Time point for use</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Enrichment of AD cases</td>
<td>CSF biomarkers may be valuable in clinical trials on early AD or MCI, to improve the diagnostic accuracy and enrich the patient sample with genuine AD cases</td>
<td>Baseline evaluation of CSF biomarkers in cases eligible for the trial</td>
<td>High T-tau and P-tau and low (\mathrm{A}\beta 42) are indicative of AD</td>
</tr>
<tr>
<td>Post-hoc patient stratification</td>
<td>AD cases with biomarker evidence of a clear disturbance in the (\mathrm{A}\beta) metabolism may have a more clear-cut effect of anti-(\mathrm{A}\beta) disease-modifying drugs</td>
<td>Post-hoc stratification of AD cases based on baseline CSF biomarker data</td>
<td>CSF (\mathrm{A}\beta 42) may be valuable to stratify AD cases enrolled in a trial on an anti-(\mathrm{A}\beta) disease-modifying drug candidate</td>
</tr>
<tr>
<td>Safety monitoring</td>
<td>CSF biomarkers may, together with MRI scans, be used to identify cases with meningoencephalitis or vasogenic edema in (\mathrm{A}\beta) immunotherapy clinical trials</td>
<td>Baseline evaluation CSF biomarkers to allow comparison with a CSF sample taken in the case of an adverse event</td>
<td>CSF cell count, IgG/IgM index, and IgG/IgM oligoclonal bands are standard measures to identify and monitor an inflammatory process, such as meningoencephalitis, within the CNS</td>
</tr>
<tr>
<td>Monitoring of drug activity on pathogenic processes</td>
<td>CSF biomarkers may provide information that the drug has an effect on a specific pathogenic process directly in patients with AD</td>
<td>Evaluation of CSF biomarkers in samples taken at baseline compared with samples taken at time points during the trial, including the last week of the trial</td>
<td>Primary CSF biomarkers for APP/(\mathrm{A}\beta) metabolism (e.g., (\mathrm{A}\beta 42), sAPP(\beta), BACE1 activity, and (\mathrm{A}\beta) turnover) may give biochemical evidence for the specific effect of an anti-(\mathrm{A}\beta) drug candidate. Downstream CSF biomarkers (e.g., P-tau and T-tau) may give biochemical evidence for downstream effects on tangle pathology and axonal degeneration of an anti-(\mathrm{A}\beta) drug candidate</td>
</tr>
</tbody>
</table>

Abbreviations: AD, Alzheimer disease; APP, amyloid precursor protein; CSF, cerebrospinal fluid; MCI, mild cognitive impairment; T-tau, total tau; P-tau, phosphorylated tau.
inflammatory CNS disorders that may mimic AD, such as neuroborreliosis (Andreason et al. 2010). The inclusion of such cases in a clinical trial might result in the erroneous conclusion that a side-effect, such as encephalitis, has occurred. Further, if a baseline CSF sample is taken for comparison before active therapy is initiated, it can be used to identify even minor inflammatory activation within the CNS that is due to side-effects of the drug, thus permitting safety monitoring in clinical trials. Lastly, biomarkers may also be valuable in demonstrating the absence of side-effects, such as immune activation, in longitudinal CSF samples during treatment.

Monitoring the Biochemical Effect of a Drug

The effects of disease-modifying anti-Aβ drugs on plaque pathology are commonly evaluated in AD transgenic mice, but these animal models have had a low predictive power for treatment success in patients with sporadic AD (Blennow et al. 2006). To bridge the gap between animal studies and large clinical trials, evidence for a true effect on AD pathogenesis directly in man would help in selecting the most promising drug candidates.

In slowly progressive disorders such as AD, evaluation of the clinical effect of a drug using rating scales requires large patient materials and extended treatment periods. For drugs with a symptomatic effect, such as cholinesterase inhibitors, an early improvement in cognitive function is expected. In contrast, a disease-modifying drug cannot be expected to have an early effect on symptoms, but will instead lead to a less pronounced decline in cognitive function over years. Thus, the number of patients needed to detect a disease-modifying effect on cognition is probably larger, and the treatment period longer, than for a symptom-modifying drug.

Biomarkers used to identify and monitor the biochemical effect of drugs are known as “theragnostic markers” (Blennow et al. 2010). Such markers may be used to identify and monitor both the specific effect of a drug on its intended target and its effect on downstream pathogenic events. A trial that uses such markers would probably require relatively small amounts of patient materials and short treatment periods. Such a strategy may be particularly suitable for making a go/no-go decision for large and expensive phase II or III clinical trials. This approach is feasible given results from longitudinal studies demonstrating low intra-individual variability of CSF T-tau, P-tau, and Aβ42 levels over time (Blennow et al. 2007; Zetterberg et al. 2007). Some of these biomarkers might also serve as substitutes (proxies) for clinical endpoints. However, full-scale clinical trials will be required to determine whether this is possible. Lastly, for regulatory purposes, a claim for a disease-modifying effect can only be made when a drug has been proven to have both an effect on cognition and biomarker evidence of an effect on the central pathogenic processes (Siemers 2009; Vellas 2009). To date, there are only preliminary reports suggesting that CSF biomarkers may be useful as theragnostic markers. Importantly, drug candidates with no proven effect on the molecular pathogenesis of AD, such as cholinesterase inhibitors and lithium, have no effect on AD CSF core biomarkers (Blennow et al. 2007; Hampel et al. 2009). Nevertheless, data from animal studies show that treatment with γ-secretase inhibitors results in a reduction of cortical, CSF, and plasma levels of Aβ (Lanz et al. 2004; Anderson et al. 2005). Similarly, treatment in monkeys with a BACE1 inhibitor results in a reduction in CSF Aβ42, Aβ40, and sAPPβ levels (Sankaranarayanan et al. 2009). In AD cases, it is uncertain how CSF Aβ may respond to treatment with efficacious anti-Aβ drugs. In a phase Ia study of the Aβ clearance-enhancing compound PBT2, a significant dose-dependent reduction in CSF Aβ42 levels during treatment was observed (Lannfelt et al. 2008). Results from a clinical study on the amyloid-targeting drug phenserine also suggest that CSF Aβ levels as a biomarker may be valuable for evaluating treatment effects (Kadir et al. 2008). However, in the interrupted phase Ia AN1792 trial, no significant effect was found on CSF Aβ42, despite a decrease toward normal levels of the downstream biomarker T-tau (Gilman et al. 2005). A clinical study of γ-secretase
inhibitor treatment also failed to find any effect on CSF Aβ42 levels (Fleisher et al. 2008). Nevertheless, this drug has a clear inhibitory effect on the Aβ production rate, as can be seen when evaluating its effect by measuring the isotope-labeled Aβ ratio in CSF; a clear inhibitory effect on the rate of Aβ production was observed (Bateman et al. 2009). Several other clinical trials of disease-modifying drug candidates that include biomarkers as endpoints are currently ongoing. These trials will provide further evidence of whether biomarkers will be useful as proof-of-concept tools for the mechanism of action of the drug, and as surrogate markers to predict clinical outcome.

FLUID BIOMARKERS AS ENDOPHENOTYPES IN GENETIC STUDIES

Just as CSF biomarker data from well-characterized, longitudinally followed cohorts may be used to guide diagnosis and estimate prognosis, it can also be used to identify genetic markers that are associated with AD risk. Compared with typical genetic studies of AD that rely on less precise clinical diagnoses, genetic studies based on quantitative endophenotype data can provide more power. In support of this approach, recent studies have shown that elevated CSF T-tau and P-tau levels are associated with single nucleotide polymorphisms in the MAPT gene (from which tau protein is produced; Kauwe et al. 2008). Likewise, CSF Aβ levels have been found to associate with polymorphisms in several genes (Kauwe et al. 2009). In this way, by “converting” endophenotype data derived from fluid biomarkers to novel genetic biomarkers, it may be possible to identify individuals at greater risk of developing AD and, in the near future, provide treatment options prior to the development of any AD pathology.

FUTURE PERSPECTIVES

Overlapping Pathology Influences Diagnostic Biomarker Accuracy

Currently available biomarkers are not perfect in diagnostic accuracy. However, except for technical shortcomings with the biomarkers, there are several fundamental reasons for why a 100% sensitive and specific biomarker for AD is an unreachable goal. First, most biomarker studies are based on clinically diagnosed cases, which introduces a relatively large percentage of misdiagnosis (Forman et al. 2006; Engelborghs et al. 2008). Second, a significant percentage of nondemented elderly have enough plaques and tangles to warrant a neuropathological diagnosis of AD (Snowdon 1997; Price and Morris 1999). Third, there is a large overlap in pathology between AD and other dementias, such as Lewy body dementia and vascular dementia (Jellinger 1996; Kotzbauer et al. 2001; Schneider et al. 2009). This overlap in pathology essentially precludes the possibility of finding biomarkers that have close to 100% sensitivity and specificity for AD. One way out of this catch 22-like situation might be to reconsider the terminology. Instead of using the term “AD biomarkers,” we could acknowledge that the biomarkers reflect distinct pathogenic or pathologic processes, for example, amyloid retention in the brain and degeneration of nonmyelinated cortical axons. These changes, especially in combination, are frequently seen in AD but may also be present in other neurodegenerative disorders, especially in isolation.

Combination of Multiple Biomarker Modalities

It is logical to suppose that the combination of CSF biomarkers with both structural (CT/MRI) and functional (SPECT/PET) brain imaging will increase the diagnostic accuracy as compared with the use of one biomarker alone. However, only a few studies to date have directly examined this issue. Positive CSF biomarkers combined with either CT or MRI measurements of medial temporal lobe atrophy have been found to increase the accuracy of an AD diagnosis (Schoonenboom et al. 2008; Zhang et al. 2008; Brys et al. 2009b). A recent study also showed that the combination of positive CSF biomarkers with the degree of structural AD-like abnormalities as shown by MRI improved prediction of the conversion from amnestic MCI to AD better than either biomarker alone.
Similarly, combining positive CSF biomarkers with an evaluation of regional cerebral blood flow using either the 133Xe method or SPECT has been shown to improve the accuracy of a prodromal AD diagnosis above that achieved with either biomarker alone (Okamura et al. 2002; Hansson et al. 2007b). Further, although no study has examined the added diagnostic value of PIB-PET and CSF biomarkers, there is a strict negative correlation between the degree of PIB binding as seen in PET images and the CSF level of Aβ42 (Fagan et al. 2006, 2009b; Forsberg et al. 2008; Grimmer et al. 2009). The relationship between PIB binding and the tau/Aβ42 and ptau/Aβ ratios is even stronger than for Aβ42 alone (Fagan et al. 2011). Large multicenter studies are needed to further define the added diagnostic value of combining multiple biomarker modalities. Such studies will also provide information on the optimal brain region to evaluate for atrophy by MRI or Aβ load by PET. Complementary data are also needed to evaluate whether new high-resolution MRI scanners and newly developed amyloid ligands, such as AZD2184 and AV-45, will improve diagnostic sensitivity and specificity (Choi et al. 2009; Nyberg et al. 2009).

Novel Research Criteria for AD
The current clinical diagnostic criteria for AD were outlined more than 25 years ago by the National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer Disease and Related Disorders (NINCDS-ADRDA) Work Group. They depend largely on the exclusion of causes other than AD for dementia (McKhann et al. 1984). These criteria state that a diagnosis of AD cannot be made until the patient has dementia, which is defined as “cognitive symptoms severe enough to interfere with social or occupational activities.” The DSM-IV and ICD-10 criteria, which are used for routine diagnosis, also require that a patient demonstrate dementia before a diagnosis of AD is possible (WHO 1992; American Psychiatric Association 2000). If the new disease-modifying drugs prove to be effective and become clinically available, these present criteria will hinder patients in the early stages of the disease, certainly in the preclinical stage, from receiving effective therapy.

For this reason, new criteria for different stages of AD have recently been suggested (Dubois et al. 2007; Albert et al. 2011; McKhann et al. 2011; Sperling et al. 2011). These criteria have been constructed to permit a diagnosis of AD in earlier stages of the disease, and are centered on the clinical identification of episodic memory impairment together with one or more abnormal biomarkers, including MRI, PET, and CSF markers. More detailed guidelines are needed to establish how the use of biomarkers can be implemented in the diagnostic procedure for early AD to be used in clinical practice. For example, details are needed regarding the scale that should be applied to measure memory impairment, which assays and cutoffs to use for CSF biomarkers, which brain region (whole brain, hippocampus, or entorhinal cortex) to use to evaluate brain atrophy by MRI, which amyloid ligand to use, and which brain region to use to evaluate brain Aβ load by PET. Studies on these issues are just beginning to emerge (Frissoni et al. 2009). Biomarker assays also need to be standardized between laboratories and centers to allow for general implementation of cutoff points in the diagnostic algorithms. As a first step in this direction, a global quality control program for CSF biomarkers was recently launched. This program also covers practical details on lumbar puncture and CSF sample processing (Blennow et al. 2010). Results from the first two rounds in this quality control program have recently been presented.

CONCLUSION
There is an enormous amount of literature showing good or excellent diagnostic performance of several biomarkers reflecting different facets of the disease process in AD. We have unprecedented possibilities to be able to phenotype our patients. Now is the time to develop the biomarker-based research criteria proposed by Dubois et al. (2007), into a detailed, practical
and feasible diagnostic algorithm that will be applicable in clinics worldwide. It is easy to predict that this will be a challenging process. The proposed algorithm would need evaluation in a longitudinal clinical multicenter study to assess its diagnostic accuracy against postconversion clinical dementia diagnoses and, whenever possible, neuropathological findings before general implementation in the clinic.

REFERENCES


Cite this article as Cold Spring Harb Perspect Med 2012;2:a006221.
Fluid Biomarkers in Medicine


Fluid Biomarkers in Alzheimer Disease


Fluid Biomarkers in Alzheimer Disease

Kaj Blennow, Henrik Zetterberg and Anne M. Fagan

Cold Spring Harb Perspect Med 2012; doi: 10.1101/cshperspect.a006221 originally published online April 17, 2012

Subject Collection The Biology of Alzheimer Disease

Animal Models of Alzheimer Disease
Frank M. LaFerla and Kim N. Green

Neurovascular Dysfunction and Faulty Amyloid β-Peptide Clearance in Alzheimer Disease
Abhay P. Sagare, Robert D. Bell and Berislav V. Zlokovic

Treatment Strategies Targeting Amyloid β-Protein
Dale Schenk, Guriqbal S. Basi and Menelas N. Pangalos

The Ubiquitin–Proteasome System and the Autophagic–Lysosomal System in Alzheimer Disease
Yasuo Ihara, Maho Morishima-Kawashima and Ralph Nixon

Neurotoxicity of Amyloid β-Protein: Synaptic and Network Dysfunction
Lennart Mucke and Dennis J. Selkoe

Proteolytic Degradation of Amyloid β-Protein
Takaomi Saido and Malcolm A. Leissring

Brain Imaging in Alzheimer Disease
Keith A. Johnson, Nick C. Fox, Reisa A. Sperling, et al.

Symptomatic and Nonamyloid/Tau Based Pharmacologic Treatment for Alzheimer Disease
Paul S. Aisen, Jeffrey Cummings and Lon S. Schneider

Alzheimer Disease in 2020
David M. Holtzman, Eckhard Mandelkow and Dennis J. Selkoe

The Genetics of Alzheimer Disease
Rudolph E. Tanzi

Fluid Biomarkers in Alzheimer Disease
Kaj Blennow, Henrik Zetterberg and Anne M. Fagan

Epidemiology of Alzheimer Disease
Richard Mayeux and Yaakov Stern

Biochemistry and Cell Biology of Tau Protein in Neurofibrillary Degeneration
Eva-Maria Mandelkow and Eckhard Mandelkow

Biochemistry of Amyloid β-Protein and Amyloid Deposits in Alzheimer Disease
Colin L. Masters and Dennis J. Selkoe

The Neuropsychological Profile of Alzheimer Disease
Sandra Weintraub, Alissa H. Wicklund and David P. Salmon

Apolipoprotein E and Apolipoprotein E Receptors: Normal Biology and Roles in Alzheimer Disease
David M. Holtzman, Joachim Herz and Guojun Bu

For additional articles in this collection, see http://perspectivesinmedicine.cshlp.org/cgi/collection/