Alcohol addiction is one of the most common and devastating diseases in the world. Given the tremendous heterogeneity of alcohol-addicted individuals, it is unlikely that one medication will help nearly all patients. Thus, there is a clear need to develop predictors of response to existing medications. Naltrexone is a $\mu$-opioid receptor antagonist, which has been approved in the United States for treatment of alcohol addiction since 1994. It has limited efficacy, in part because of noncompliance, but many patients do not respond despite high levels of compliance. There are reports that a missense single nucleotide polymorphism (rs179919 or A118G) in the $\mu$-opioid receptor gene predicts a favorable response to naltrexone if an individual carries a “G” allele. This work will review the evidence for this hypothesis. The data are promising that the “G” allele predisposes to a beneficial naltrexone response among alcohol-addicted persons, but additional research is needed to prove this hypothesis in prospective clinical trials.

Ventral tegmental neurons release dopamine at nerve terminals in ventral striatum and medial prefrontal cortex. Activation of this circuit is a common element of abused drugs, including alcohol (e.g., Di Chiara and Imperato 1988; for review, see Koob and Volkow 2010). Thus, alcohol shares in common with nicotine, cocaine, amphetamine, morphine, etc, this property of enhancing dopamnergic transmission in ventral striatum and medial prefrontal cortex. Both animal model and human studies are in agreement on this point (Boileau et al. 2003; Gilman et al. 2008; Spanagel 2009). This release of dopamine in the ventral striatum and medial prefrontal cortex is partially enhanced by stimulation of $\mu$-opioid receptors (for which endorphin is the primary ligand) located on inhibitory GABAergic interneurons in the ventral tegmental area. The GABAergic interneurons inhibit the dopamnergic ventral tegmental neurons, whose activation signals reward. Thus, $\mu$-opioid receptor agonists enhance the likelihood of ventral tegmental dopaminergic neuron activation (and the experience of reward) by lessening the tonic inhibition of the associated GABAergic interneurons (Johnson and North 1992; Spanagel et al. 1992; Tanda and DiChiara 1998).

Given this circuitry, it has been consistently shown that endogenous opioids play a role in ethanol reinforcement in various animal paradigms. Endorphin elevations after alcohol are seen in discrete reward regions of the hypothalamus (Popp and Erickson 1998), ventral tegmentum, and ventral striatum (Rasmussen et al. 1998). It is important to note that endorphin deficient rats continue to self-administer
alcohol, indicating that endorphin is not the sole mechanism of alcohol reward (Grahame et al. 1998). The importance of \( \mu \)-opioid receptor activation as a mechanism for alcohol reward is underscored by the fact that alcohol consumption in alcohol-preferring rats is persistently reduced after inactivating \( \mu \)-opioid receptors in the ventral striatum (Myers and Robinson 1999). Similarly, decreased alcohol self-administration is observed in primates after pretreatment with opioid antagonists (Altshuler et al. 1980). C57Bl/6J mice, an inbred strain that prefers alcohol, have increased endorphin release in the hypothalamus after alcohol administration (De Waele et al. 1992). Alcohol preferring rats have high levels of opioid gene mRNA species in the hypothalamus, prefrontal cortex, and mediodorsal nucleus of the thalamus (Marinelli et al. 2000), as well as increased \( \mu \)-opioid receptor density in the ventral striatum and medial prefrontal cortex.

**CLINICAL STUDIES OF NALTREXONE IN ALCOHOLISM**

The development of a substantial body of evidence, in the 1980s, that naltrexone (an orally active \( \mu \)-opioid receptor antagonist) diminished alcohol self-administration in animal models (Altshuler et al. 1980; Kiianmaa et al. 1983; Myers et al. 1986; Volpicelli et al. 1986) led to the first use of naltrexone in alcohol addicted populations in a controlled clinical trial (Volpicelli et al. 1992), the promising outcome of which was immediately confirmed in a second controlled clinical trial (O’Malley et al. 1992). Naltrexone was found to reduce alcohol craving and relapse to heavy drinking (operationally defined as five or more drinks/day for a man, four or more for a woman), but did not reduce abstinence rates. On the basis of these two controlled trials, naltrexone was approval by the FDA, in the absence of the usual pharmaceutical industry interest.

In the intervening 20 years, there have been more than 30 clinical trials of naltrexone in alcohol addiction (for review, see Bouza et al. 2004; Srisurapanont and Jarusuraisin 2005; Pettinati et al. 2006). Although the majority of these clinical trials show efficacy of naltrexone in reducing risk for relapse to heavy drinking, the effect size is small, with many patients having no benefit. This has resulted in multiple reports in which the naltrexone arm outcomes are not significantly better than the placebo arm outcomes (e.g., Krystal et al. 2001). This is an expected outcome, given the tremendous heterogeneity of clinical alcohol addiction. It is likely that important clinical characteristics, such as compliance, severity and duration of alcohol addiction, comorbidity (both medical and psychiatric), and/or attendance at psychosocial treatment, may influence outcomes.

In this situation, multiple investigators have attempted to define clinical characteristics, which might enhance the probability of naltrexone response. Some clinical measures have shown promise in characterizing a naltrexone responder: high alcohol craving (Chick et al. 2000; Monterosso et al. 2001; O’Malley et al. 2002) and strong family history of alcohol addiction (Monterosso et al. 2001), but family history of alcohol addiction did not predict response to naltrexone in the COMBINE multicenter trial (Capone et al. 2011). Alcohol addicts who experience greater euphoria after alcohol may have a better response to naltrexone (Volpicelli et al. 1995).

**A118G OPRM1 MISSENSE SINGLE NUCLEOTIDE POLYMORPHISM: MOLECULAR AND CELLULAR EFFECTS**

A common missense single nucleotide polymorphism (rs 1799971) in the first exon of the \( \mu \)-opioid receptor gene, OPRM1, was described by Bergen et al. (1997), A118G, or N40G, reflecting the fact that the A allele encodes asparagine, whereas the minor G allele encodes aspartate. The A (asparagine) allele is thought to be N-glycosylated (Huang et al. 2012), whereas this is not possible for the G (aspartate) allele, as there is no free amino group. Subsequent study (e.g., Gelernter et al. 1999; Szeto et al. 2001; Crowley et al. 2003; Tan et al. 2003) revealed large ethnic differences in allele frequencies (see Table 1).

This allele has been the subject of multiple molecular investigations to determine its
functional consequences, in terms of gene expression, protein translation, receptor signaling, and receptor density. Initially, Bond et al. (1998) reported that the minor “G” allele μ-opioid receptor resulted in decreased affinity for binding to β-endorphin, compared to the common “A” allele receptor. There was no change in binding affinity for alkaloid ligands. This result has not been confirmed in subsequent investigations (Beyer et al. 2004; Ramchandani et al. 2010). In one such study transfected HEK293 cells (a fibroblastoid cell type) were used (Beyer et al. 2004), but the 118G allele did not differ in binding affinity for β-endorphin, compared to 118A. Beyer et al. (2004) also reported that the 118G allele was not different from the 118A allele in rate of desensitization, internalization or resensitization, but 118G had decreased transcription, compared to 118A. Ramchandani et al. (2010) also did not report differences in kinetics of binding of β-endorphin to the 118G, compared to 118A. Mahmoud et al. (2011), using a whole cell patch clamp technique in acutely dissociated trigeminal ganglion neurons, reported that morphine was fivefold less active at the “G” allele receptor form in activating a Ca²⁺ channel. There was no such difference for fentanyl. Zhang et al. (2005) conducted allelic imbalance studies in postmortem human brain, revealing a marked decrease in 118G allele mRNA (see Fig. 1). In a second experiment, they showed in vitro evidence of a marked decreased translation of the 118G mRNA (see figures in Ramchandani et al. 2010) (Zhang et al. 2005).

### Table 1. Frequency of G allele for A118G SNP in ethnic groups

<table>
<thead>
<tr>
<th>Ethnic group</th>
<th>Freq G</th>
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<th>Freq G</th>
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<tr>
<td>African</td>
<td>1%</td>
<td>Korean</td>
<td>31%</td>
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<tr>
<td>African-American</td>
<td>3%</td>
<td>Chinese</td>
<td>35%</td>
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<tr>
<td>Swedish</td>
<td>11%</td>
<td>Malaysian</td>
<td>43%</td>
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<tr>
<td>European-American</td>
<td>15%</td>
<td>Indian</td>
<td>47%</td>
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A118G OPRM1 MISSENSE SINGLE NUCLEOTIDE POLYMORPHISM: ANIMAL MODEL STUDIES

In the murine OPRM1 gene, there is no equivalent of the A118G naturally occurring variation. A homologous variation (A112G, with the A allele encoding asparagines and the G allele encoding aspartate, as in the human OPRM1 gene) was created by bacterial artificial chromosome engineering and murine transgenic techniques by Mague et al. (2009). They reported decreased transcription and translation of the G allele in transgenic C57Bl/6 mouse brain (see Fig. 2, a result congruous with the human postmortem brain ex vivo results of Zhang et al. 2005), as well as in vitro results of Beyer et al. (2004). There was a blunted locomotor response to morphine in the 112G mice, as well as decreased morphine

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![Figure 1](http://perspectivesinmedicine.cshlp.org/)

**Figure 1.** Transcriptional and translational efficiency of the 118G allele is markedly limited, compared to the A allele. Allelic imbalance was shown in the human postmortem brain, with the 118G showing marked transcriptional decrease. Translation of the 118G was also decreased using an in vitro approach. *p < 0.05; **p < 0.01. (Created from data by Zhang et al. 2005.)
conditioned place preference (CPP) in 112G female mice, the latter being a sexually dimorphic response, with 112G males showing the expected CPP response to morphine.

Two other forms of transgenic mice were produced, using homologous recombination to replace the murine OPRM1 exon 1 with one of the two forms (118A and 118G) of human OPRM1 exon 1 (Ramchandani et al. 2010). These investigators conducted in vivo microdialysis experiments in the ventral striatum, demonstrating that the 118G mice had the expected elevations in dopamine release after alcohol, whereas the 118A mice had no significant increase over baseline (see AA and AG group figures in Ramchandani et al. 2011). These data suggest that the “G” allele conveys an increased rewarding valence to alcohol, compared to the “A” allele.

There have been several studies of a similar SNP in the rhesus monkey, the C77G, which results in a homologous amino acid change, asparagine to aspartate (Fig. 3) (Barr et al. 2007, 2010; Vallender et al. 2010). Both groups report that the G allele monkeys consume significantly more alcohol than the CC monkeys. Further, both groups note that naltrexone significantly decreases alcohol intake in the GG monkeys.

These reports, taken together, are consistent with the hypothesis that the 118G allele (or its equivalent in mouse and primate) conveys a greater rewarding effect of alcohol, a difference that is inhibited by naltrexone. These studies are remarkably consistent, given the species, paradigm, technical, and molecular engineering differences among these studies.

A118G OPRM1 MISSENSE SINGLE NUCLEOTIDE POLYMORPHISM: HUMAN PHARMACOGENETIC STUDIES OF ALCOHOL

There have been several pharmacogenetic reports of the A118G SNP in human laboratory...
experiments involving alcohol (Ray and Hutchison 2004, 2007; Ramchandani et al. 2010; Ray et al. 2010; Setiawan et al. 2011). In a laboratory investigation of the A118G pharmacogenetics of alcohol reward, Ray and Hutchison (2004, 2007) showed that the G allele carriers experienced significantly greater euphoria after standard oral doses of alcohol (while controlling for breath alcohol concentration), compared to AA persons. Further, naltrexone significantly blunted the euphoria in the G allele carriers and was without effect in the AA group (see Fig. 4).

In agreement with this result, Ramchandani reported that G allele carriers had a greater striatal release of dopamine after alcohol (using a raclopride PET scan technique), compared to AA participants (see Ramchandani et al. 2011). In a more naturalistic approach, Ray et al. (2010) studied drinking habits of social drinkers over a 5-d period, analyzing subjective responses to alcohol by A118G genotype. G allele carriers reported more significantly more “vigor” less negative mood after drinking, compared to the AA group. Similarly, Setiawan et al. (2011) studied the subjective response to alcohol in social drinkers after a dose of naltrexone. Naltrexone significantly decreased the ethanol-induced “euphoria” to a priming dose of alcohol in subjects with the G allele, compared to AA participants.

Taken together, these human laboratory studies of the A118G variant on effect of alcohol are remarkably consistent, with the clear conclusion that the G allele permits people to experience alcohol in a more rewarding manner, compared to AA individuals. It is also notable that naltrexone is able to blunt this euphoria in G allele carriers, but not in AA persons. This latter observation is consistent with subjective reports of the effect of naltrexone in clinical trials for alcohol addiction, in which the medication attenuated alcohol-induced euphoria among responders (Volpicelli et al. 1995).

Figure 4. (A) 118G allele carriers have a greater euphoria response to alcohol, compared to homozygous A persons; (B) naltrexone blunts the alcohol-induced euphoria for 118G carriers, but has no such effect in persons who are homozygous A. (Created from data by Ray and Hutchinson 2007.)
retrospective analysis of three naltrexone trials of similar design, two conducted at the University of Pennsylvania and one at the University of Connecticut. Compliance was monitored by riboflavin testing and by pill counts. Eighty-two patients (71 of European descent) who were randomized to naltrexone and 59 randomized to placebo (all of European descent) in one of three randomized placebo-controlled clinical trials of naltrexone were genotyped at the A_{118}G (Asn40Asp) and C_{17}T (Ala6Val) SNPs in the μ-opioid gene (OPRM1). The association between genotype and drinking outcomes was measured over 12 wk of treatment. For purposes of examining the pharmacogenetics of naltrexone response, the analysis was limited to those subjects with well-defined outcome data who had at minimum 6-wk exposure to the medication. The primary drinking outcome considered was relapse to heavy drinking (≥5 drinks in a single day for men or ≥4 drinks for women). This definition of heavy drinking was the primary outcome for each of the trials. The time line follow back method was employed (along with self report) to measure alcohol consumption (Sobell and Sobell 1992). There was a significantly greater proportion of naltrexone treated subjects with the G allele variant who did not return to heavy drinking (no relapse) compared to those with those homozygous for the A allele (Wald = 4.04, 1 df, OR = 3.47 [95% CI: 1.03–11.67], \( p = 0.045 \); see Table 2).

This finding was confirmed in a larger multisite study of naltrexone, acamprosate, and placebo for alcohol addiction (Anton et al. 2008). Alcohol addicted subjects were treated for 16 wk with 100 mg of naltrexone. All participants received medical management alone or with combined behavioral intervention. The combined behavioral intervention was delivered by licensed behavioral health specialists in up to 20 flexible participant need-adjusted 50-min sessions. Combined behavioral intervention, an intensive and specific alcohol intervention, may have compensated for the placebo effect, thereby suppressing the chances of observing a main effect of naltrexone or a genetic interaction. The data presented by Anton et al. (2008) are consistent with this thinking. A gene X medication interaction may be observable only in patients who can show obvious benefit from the medication over placebo.

In a small Korean study of naltrexone in alcohol addiction (Kim et al. 2010), subjects adherent to naltrexone treatment with one or two copies of the Asp40 allele took a significantly longer time than the Asn40 group to relapse to heavy drinking (\( p = 0.014 \)). Although not significant, the Asn40 group treated with naltrexone had a 10.6 times greater relapse rate than the Asp40 variant group. There was no effect on abstinence.

When considering only those patients receiving medical management alone, there was an significant effect of naltrexone on “good outcome” among the 118G carriers, whereas there was no such effect for the patients receiving naltrexone who were homozygous A118 (see Table 2). However, there was no such effect in the naltrexone group receiving medical management with combined behavioral intervention. The combined behavioral intervention was delivered by licensed behavioral health specialists in up to 20 flexible participant need-adjusted 50-min sessions. Combined behavioral intervention, an intensive and specific alcohol intervention, may have compensated for the placebo effect, thereby suppressing the chances of observing a main effect of naltrexone or a genetic interaction. The data presented by Anton et al. (2008) are consistent with this thinking. A gene X medication interaction may be observable only in patients who can show obvious benefit from the medication over placebo.

In the Veterans Administration multisite study of naltrexone in alcohol addiction, Geiernter et al. (2007) reported that the 118G allele did not predict outcome among 149 participants in the naltrexone group and 64 in the placebo group. There are several possible explanations for this result. First, the efficacy of

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<th>Table 2. A118G genotype and good outcome in naltrexone studies of pharmacotherapy for alcohol addiction</th>
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<td>Genotype at A118G</td>
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<tr>
<td>G allele carriers</td>
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<tr>
<td>Homozygous A</td>
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<tr>
<td>Placebo</td>
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<sup>a</sup>\( P = 0.04, \text{ odds ratio} = 3.5 \).
<sup>b</sup>\( P = 0.005, \text{ genotype} \times \text{ medication interaction; odds ratio} = 5.8 \).
naltrexone is certainly influenced by compliance, and the compliant population was defined as those who opened the medication bottle a minimum of 50% of the time, so that medication compliance was defined liberally. Second, it is likely that high levels of comorbidity influence response to naltrexone. The study population had substantial rates of recurrent unipolar illness, antisocial personality and anxiety disorders, and had severe alcohol addiction of long duration. These factors might overwhelm any genetic predisposition to respond to naltrexone. Third, the study had limited power; for example, there were only nine 118G carriers in the placebo group.

Coller et al. (2011) recently reported the results of a naltrexone and cognitive-behavioral therapy trial in 100 Australian alcohol addicted persons. They reported an overall effect of naltrexone on relapse to heavy drinking, but no influence of the A188G variants. The absence of a control group makes this study less ideal, as does the small sample size, with 68 study completers.

Taken together, the A118G clinical trials in naltrexone treatment for alcohol addiction remain promising, but there are clear unanswered questions, including the influence of counseling, compliance, and comorbidity on outcome. Available depot formulations of naltrexone may reduce noncompliance, but the influence of comorbidity and counseling may be more difficult to resolve. It will be necessary to conduct pharmacogenetic alcohol addiction naltrexone trials, for which participants are randomized by A118G genotype into the naltrexone or placebo arm to reduce possible sources of bias. These trials should be characterized by:

1. large size (at least 150 persons per arm, including oversampling of G allele carriers) to ensure adequate power;
2. rigorous assessment of compliance;
3. randomization stratified by genotype;
4. careful assessment of comorbidity; and
5. modest psychotherapeutic intervention, so as to mirror “real world” clinical practice.

**SUMMARY**

There are extensive data, across species, to suggest that the 118G form of the μ-opioid receptor is characterized by decreased transcription and translation. There are convincing data, from murine, primate, and human laboratory studies, that the 118G (or its species-specific homolog) variant permits alcohol to have a greater rewarding valence, leading to increased alcohol consumption. Further, the human and rhesus data are equally convincing that naltrexone is able to blunt this greater rewarding signal. Last, the possibility that A118G alleles can be used clinically to identify alcohol addicted persons with a greater probability to have a beneficial response to naltrexone is a hypothesis that deserves testing on a large scale, with the characteristics noted above.

**REFERENCES**


Pharmacogenetics of Alcohol Addiction


Opioid Pharmacogenetics of Alcohol Addiction

Wade Berrettini

Cold Spring Harb Perspect Med 2013; doi: 10.1101/cshperspect.a012203 originally published online May 31, 2013

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